Analysis of the evolution in the circulation of HAV and HEV in Eastern Spain by testing urban sewage samples

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ABSTRACT

The aim of the study was to analyse the evolution of the prevalence of HAV and HEV in the population of eastern Spain by analysing the viruses excreted in urban sewage. Raw urban sewage samples were collected and analysed during several years using RT-PCR techniques and sequencing analysis. Two limiting regions were analysed, one of them having implemented HAV vaccination programs. Acute symptomatic HEV cases were also examined. Results were compared with those from previous studies in the area using identical methodology. The percentage of positive HAV samples in urban sewage fell from 57.4% to 3.1% in 5–10 years in the two studied areas in Spain. Around 30% of the urban sewage samples were positive for HEV in the absence of agricultural sources of contamination. HEV RNA was also detected in four clinical cases of acute hepatitis. The dramatic reduction in the presence of HAV in raw urban sewage observed in eastern Spain could be most likely related to the general improvement in sanitation. However, these improvements would not have an equivalent effect on the circulation of HEV and this observation could be explained by the presence of animal reservoirs for HEV, which act as external sources of infections.

Key words | acute hepatitis, HAV, HEV, sanitation, vaccination, wastewater

ABBREVIATIONS

BLAST | Basic Local Alignment Search Tool
DNA | Deoxyribonucleic acid
EIA | Enzyme immunoassay
EMEM | Eagle’s minimum essential medium
FDA | Food and Drug Administration
GC | Genomic copies
HAdV | Human adenovirus
HAV | Hepatitis A virus
HBeAg | Hepatitis B e-antigen
HBsAg | Hepatitis B surface antigen
HCV | Hepatitis C virus
HEV | Hepatitis E virus
IgG | Immunoglobulin G
IgM | Immunoglobulin M
NAs | Nucleic acids
NCBI | National Center for Biotechnology
NE | Northeast
NJ | Neighbour joining

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INTRODUCTION

Hepatitis A virus (HAV) and hepatitis E virus (HEV) are small, non-enveloped viruses that contain positive sense, single-stranded RNA genomes of 7.5 and 7.2 kb, respectively (Purcell & Emerson 2008). HAV has its own genus (Hepatovirus) within the family Picornaviridae whereas HEV is the sole member of the genus Hepevirus and family Hepeviridae (Emerson et al. 2005; Nainan et al. 2006). Both viruses cause acute, self-limited infections that may vary in severity from asymptomatic to fulminant (Purcell & Emerson 2008). The most common route of transmission is generally faecal-oral (Orru et al. 2004) and it causes large-scale epidemics.

Hepatitis A affects humans worldwide. HAV infection is endemic to developing countries where most individuals are exposed to this virus during childhood (Jacobsen & Koopman 2004). Hepatitis E is the major cause of waterborne outbreaks and sporadic cases of acute hepatitis in many developing countries where sanitation is suboptimal and where HEV is considered endemic (Clemente-Casares et al. 2003; Emerson & Purcell 2004). Typically, the disease caused is self-limited but it sometimes has severe complications, particularly in pregnant women, who register a high mortality rate (20%) (Balayan 1997).

Traditionally, North America and Europe have been considered non-endemic regions for HEV as they have seroprevalence rates of 1 to 5% and most HEV infections are thought to be imported (Paul et al. 1994). However, in the last few years several HEV strains associated with sporadic acute hepatitis and genetically distinct from the endemic ones have been isolated from human serum samples in North America and in some European countries (Clemente-Casares et al. 2009; Legrand-Abravanel et al. 2009). A recent study conducted in the same area, Catalonia (NE Spain), showed a seroprevalence of about 7.3% (Buti et al. 2006). There is also evidence that some animals, particularly swine, are infected by HEV genotypes 3 and 4. Both genotypes have been recovered from swine principally in the same regions that these genotypes were recovered from humans (Meng et al. 1997; Purcell & Emerson 2008). With few exceptions, genotypes 3 and 4 appear not to produce disease in swine (Martín et al. 2007). In certain regions, including Spain, swine and human HEV strains are not genetically distinguishable (Clemente-Casares et al. 2009), thereby supporting the notion that swine are a reservoir and that HEV infection is acquired by handling these animals or eating raw pork products.

Spain is considered an area of low endemicity for HAV infection (Bell 2002) and is characterized by diminishing HAV seroprevalence in the population (Jacobsen & Koopman 2004), while it is a non-endemic area for HEV (Yarbough 1999). Previous studies in Spain show that even in the absence of significant numbers of clinical cases of acute hepatitis A and E, HAV and HEV are present in more than 50 and 40%, respectively, of urban sewage samples analysed, thereby indicating a high level of subclinical infection in the population (Pina et al. 2001; Clemente-Casares et al. 2005).

In 2001, the FDA approved a new combination vaccine against HAV and the hepatitis B virus. The vaccine, called Twinrix, contains the approved vaccine Havrix® (Hepatitis A Vaccine, Inactivated) (US Food and Drug Administration 2001). Twinrix has been distributed in a pilot program in Catalonia for pre-adolescents since 1999. In Valencia, a region immediately south of Catalonia, HAV vaccines are distributed only among groups at risk as a result of handling food. No HEV vaccine is currently available for humans.

By 2006, 255,170 and 11,826 doses of vaccines against HAV had been distributed in Catalonia (covering around 90% of pre-adolescents) and Valencia (in the professional food handler group), respectively (Departament de Salut, Generalitat de Catalunya 2008; Direcció general de salut pública, conselleria de sanitat, Generalitat Valenciana 2008).

This study analyses changes in the excretion patterns of HAV and HEV in eastern Spain by testing the presence of these viruses in urban sewage over several years in order to evaluate the potential impact of improvements in sanitation.
and vaccination programs for HAV. Serum samples from acute hepatitis patients were also analysed to confirm the presence of HEV strains in the population.

**METHODS**

**Sewage samples**

Sampling areas were selected on the basis of the vaccination programs applied for HAV. Samples were collected in Barcelona and Valencia, the largest cities in Catalonia and Valencia respectively, both in Spain. A total of 91 urban sewage samples were collected in Barcelona (50 samples) and Valencia (41 samples) from 2000 to 2008. Samples were collected at the entry of two treatment plants in Catalonia receiving sewage from a population of about 1.8 million (Barcelona) and three plants from the area of Valencia, with a total population of about 800,000 inhabitants. Each sample was harvested in a sterile 500-ml polyethylene container and kept at 4°C for less than 8 h until the virus particles were concentrated.

**Clinical samples**

From 2004 to 2007, 19 serum samples were collected from patients with symptomatic acute hepatitis attending the Hospital General Valle Hebron (Barcelona, Spain). All patients had elevated aminotransferase levels (>10 times the upper normal limit) and IgG anti-HEV antibodies. Clinical serum samples were also analysed for other viral infections. Hepatitis B surface antigen and antibody to HCV (anti-HCV) were detected by commercial enzyme immunoassays (Vitros HBsAg and Vitros anti-HCV; Johnson & Johnson, Rochester, NY, USA). IgM anti-HBc and IgM anti-HAV were detected by enzyme immunoassays (Liaison HBeAg/anti-HBe, Liaison HBc IgM and Liaison HAV IgM, respectively; Diasorin, Vercelly, Italy). IgG anti-HEV was the routine assay in use for HEV infections and was determined by EIA (Bioelisa HEV IgG; Biokit, Barcelona, Spain). Serum samples were stored at −80°C until use for detecting genomic sequences. Other causes of acute hepatitis such as autoimmune hepatitis or biliary stones were excluded.

**Viruses**

The positive controls used for the analysis of HAV consisted on dilutions of the supernatant of FRhK-4 cell cultures infected with pHM-175 strain. As positive controls for HEV analysis, 10% faecal suspensions obtained from rhesus monkeys experimentally infected with a strain isolated from a sewage sample collected in Barcelona (Pina et al. 1998) were used. HAdV2, isolated from a clinical sample and used as positive control for the determinations of human adenoviruses, was grown on A549 cells propagated in Eagle's minimum essential medium (EMEM) supplemented with 1% glutamine, 50 μg of gentamicin per ml, and 5% (growth medium) or 2% (maintenance medium) of heat-inactivated FBS. Viral suspensions were stored at −80°C until use.

**Concentration of viruses from urban sewage**

The method used to recover virus particles from sewage samples was chosen on the basis of previous studies (Puig et al. 1994; Pina et al. 1998). Briefly, 42 ml of sewage was ultracentrifuged at 110,000 × g for 1 h at 4°C to pellet all the viral particles together with any suspended material. The pellet was eluted by mixing it with 3.5 ml of 0.25 N glycine buffer (pH 9.5) on ice for 30 min and, after addition of 3.5 ml of 2X phosphate-buffered saline, the suspended solids were separated by centrifugation at 12,000 × g for 20 min. Finally, viruses were concentrated by ultracentrifugation at 110,000 × g for 1 h at 4°C, resuspended in 100 μl of phosphate-buffered saline and stored at −80°C.

**Nucleic acid extraction**

The nucleic acids (NAs) from viral concentrates obtained from sewage samples were extracted following a highly sensitive and efficient procedure (Puig et al. 1994) described by Boom et al. (Boom et al. 1990). This procedure is based on the use of guanidinium isothiocyanate and silica particles.

For clinical serum samples, QIAamp RNA Blood Mini Kit® (Qiagen, Valencia, Spain) was used for the viral RNA extraction, following the manufacturer’s instructions.
Reverse transcription-polymerase chain reaction

All samples were analysed with primers designed to amplify a region within the ORF2 of HEV (Erker et al. 1999) (semi-nested RT-PCR) and a non-coding region of 5' end of HAV (Pina et al. 2001) (nested RT-PCR). The VP1/2A region was used to characterize HAV-positive samples by sequencing (Pina et al. 2001). Positive and negative controls extracted in parallel and inhibition controls were added to the PCR assays. The results of the PCRs using OneStep® RT-PCR kit from Qiagen, for a typical one-step reaction, and the 50-μl reaction mixture containing PCR Gold Buffer, 1.2 mM MgCl$_2$ and 2 U of AmpliTaq® Gold (Applied Biosystems, New Jersey, USA), for a second PCR amplification cycle, were analysed by electrophoresis on agarose gels to 3% (w/v), followed by staining with ethidium bromide at 0.5 μg/ml. The amplicons were viewed using an Image Master® VDS (Pharmacia Biotech, Uppsala, Sweden).

Analysis of sewage samples by QPCR

HAV- and HEV-positive samples by nested PCR were quantified by real-time PCR (Jothikumar et al. 2005b, 2006). Five microlitres of the extracted RNA, corresponding to 2.1 ml sewage, was analysed. Five microlitres of ten-fold dilutions (from 5 × 10$^8$ to 5 × 10$^7$ GC/5 μl) of a plasmid suspension were used as standard. Amplification reactions were performed with the QuantiTect® Probe RT-PCR kit reagents (Qiagen).

Human adenoviruses (HAdVs) were determined in each location. The viral load of HAdV was used to discard negative HAV and HEV results caused by the absence of human faecal contamination, which could influence the results of the study (Bofill-Mas et al. 2006).

QPCR amplification reactions for HAdV were carried out using TaqMan® Universal PCR Master Mix reagents (Applied Biosystems), as previously described (Jothikumar et al. 2005a; Bofill-Mas et al. 2006). The HAdV 41 used in the construction of the standards was kindly donated by Dr Annika Allard at the University of Umeå, Sweden. Ten microlitres of the NA extractions were analysed, corresponding to 4.2 ml of sewage sample. Dilutions from 10$^{-1}$ to 10$^3$ CG/10 μl of the standard were analysed in triplicate.

In all PCR assays, standard precautions were taken by using separate areas for the diverse steps of the protocols. Negative controls, inhibition controls and positive controls were added in each assay. Positive results were confirmed by sequencing analysis of the amplified DNA. The QPCRs were conducted in a thermocycler Stratagene® MX3000P (Stratagene, La Jolla, USA). All the samples and their respective dilutions were analysed in duplicate.

Sequencing and analysis of viral genomes

Amplicons obtained with the nested PCR were purified using a QIAquick® purification kit (Qiagen), following the manufacturer’s instructions. Both strands of the purified DNA amplicons were sequenced using an ABI Prism 3100 Genetic Analyzer and Big Dye Terminator Cycle Sequencing Kit v. 3.1® (Applied Biosystems), following the manufacturer’s instructions. Products were checked using an ABI PRISM 377 analyser (Applied Biosystems) by the Scientific and Technical Services of the University of Barcelona. The sequences were compared with the nucleotide sequences present in the GenBank using the BLAST program of the NCBI (National Center for Biotechnology Information; www.ncbi.nlm.nih.gov/BLAST) and aligned using the Clustal X 1.8 program. Phylogenetic tree reconstructions were performed with Mega 3.1 (www.megasoftware.net). Evolutionary distances were determined by the Kimura two-parameter equation, and the tree was constructed by using the neighbour-joining (NJ) algorithm. The GenBank accession numbers for the sequences reported in this study are FJ010829-FJ010840 (HAV in sewage), FJ010841-FJ010844 (HEV in serum samples) and FJ010845-FJ010858 and EU729700 (HEV in sewage).

RESULTS AND DISCUSSION

This study was designed as a comparative analysis for the evaluation of changes in excretion patterns of acute hepatitis viruses in eastern Spain. Previous studies conducted in our laboratory between 1994 and 2002 analysed environmental and clinical samples using the same protocols as in this study and reported a frequent presence of HAV and HEV in urban sewage (Pina et al. 2001;
The recent availability of an efficient vaccine against HAV infections, the implementation of vaccination programs for a high proportion of the susceptible population and a general improvement in sanitary conditions are expected to decrease the circulation of the virus in the population and the levels of excretion of HAV in the urban sewage generated.

Presence and concentration of HAV and HEV

Average viral concentrations in wastewater, as determined by the QPCR assays, were $1.6 \times 10^2$ GC/ml for HAV and $3.2 \times 10^1$ GC/ml for HEV. The results on the prevalence of positive HAV and HEV samples using RT-PCR assays were equivalent in the two regions in NE Spain (Table 1).

HEV was detected in clinical serum samples using nested RT-PCR. Four of the 19 analysed serum samples analysed were positive for HEV RNA, with one sample also being positive for IgM anti-HAV and one positive for anti-HCV antibodies without HCV RNA. These results confirm that HEV circulates in the population and that it causes sporadic clinical cases of acute hepatitis.

In 2001, Pina et al. (Pina et al. 2001), using the same methodology as in the present study, analysed HAV strains excreted in sewage in the area of Barcelona and also the HAV strains responsible for acute hepatitis A clinical infections in Barcelona. These authors found that 57.4% (31/54) of the sewage samples collected from 1994 to 2000 were positive for HAV and identified clinical HAV infections. In addition, the distribution of HAV was found to be equivalent to the strains detected in urban sewage. That study is considered a reference for the evaluation of changes in the epidemiological pattern of HAV in the study area.

The results obtained in this study show a marked reduction in the prevalence of HAV in the study area, thereby suggesting that the higher risk of this population to acquire HAV infection is at present through personal or food-borne transmission from other areas with a higher level of endemicity.

General immunization in adolescents has a protective effect on the population; however, the pattern of HAV excretion in urban sewage did not differ significantly between the two regions in Spain, one of which has a general vaccination program while the other does not. However, what is unquestionable is that vaccination is the only measure that guarantees individual protection against HAV infections in susceptible populations.

Improvements in sanitation would include not only improvements in water treatment but also improvements in hygienic and preventive practices that will better protect the environment, water and food from contamination, reducing the overall exposure and the number of infections and excreted viruses in the population. HAV is known to be a very stable virus (Siegl et al. 1984). The samples analysed in this study were raw urban sewage samples presenting standard concentrations of human adenoviruses and without any treatment (Bofill-Mas et al. 2006). The observed differences in the presence of HAV in the samples would be then related with higher probability to the reduction in the prevalence and excretion of hepatitis A viruses in the population than to significant changes in the stability of the viruses or in the level of faecal contamination in the sewage.

Table 1: Presence of hepatitis A virus (HAV) and hepatitis E virus (HEV) in urban wastewater

<table>
<thead>
<tr>
<th>Country/Area</th>
<th>Year</th>
<th>HAV (Nested PCR)</th>
<th>HAV (Semi-nested PCR)</th>
<th>HEV (Nested PCR)</th>
<th>HEV (Semi-nested PCR)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Spain/Barcelona</td>
<td>1994–2002</td>
<td>31/54 (57.4%)</td>
<td>20/46 (43.5%)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Spain/Barcelona</td>
<td>2000–2004</td>
<td>5/18 (27.8%)</td>
<td>7/18 (38.9%)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Spain/Barcelona</td>
<td>2006–2008</td>
<td>1/32 (3.1%)</td>
<td>9/32 (28.1%)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Spain/Valencia</td>
<td>2007–2008</td>
<td>1/41 (2.4%)</td>
<td>13/41 (31.7%)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

The results for nested and semi-nested assays are represented as positive samples/samples analysed (percentage of positives samples).

The results shown were obtained from previous studies Pina et al. (2001) and Clemente-Casares et al. (2003), respectively.

Concentration of HAdV in wastewater samples

The level of human faecal contamination of the sampling sites was evaluated by determining the concentration of HAdV using QPCR. As expected, HAdV was detected in 100% of the analysed samples and the main value of concentration was $1.5 \times 10^5$ GC/ml of water tested. These values are frequently observed in urban sewage and are used as a marker of faecal contamination in contaminated water (Bofill-Mas et al. 2006).
Genetic characterization of the HAV and HEV strains

The sequences of HAV and HEV detected were compared with sequences described in previous studies in the same area and sequences available in databases. Five out of seven positive HAV sequences detected by amplification of the VP1/2A region (290 bp) were genotyped. The strains detected in Spain belonged to genotype IA and IB. Only two urban sewage samples were positive for HAV among the samples collected in Spain between 2006 and 2008, one from Barcelona, genotype IB, and one from Valencia, genotype IA.

Fifteen out of 29 positive samples for HEV were genotyped and the HEV strains detected were analysed by comparing the 101 bp sequence amplified in the ORF2 region. As expected, most of the HEV strains detected in samples from Barcelona and Valencia corresponded to genotype 3. However, in one sample from Barcelona, a genotype 1 HEV strain was identified. Three of the HEV strains identified from serum samples belonged to genotype 3, and one to genotype 1, the latter case corresponding to a person that had travelled to India during the previous month. Phylogenetic relationships between HEV strains isolated in Barcelona and Valencia were established by comparing the representative strains (Figure 1).

CONCLUSIONS

A previous study on HEV in the area around Barcelona (Clemente-Casares et al. 2003) showed a prevalence of positive samples of 43.5% (20/46) in urban sewage samples collected during 1994–2002. In the last years the prevalence appeared to be established at around 30% in the two study areas of Spain, with average concentrations of 3.2 × 10^3 GC/ml. These results demonstrate that HEV is circulating in the human population of this area and that urban sewage may be a source of contamination by this pathogen. Moreover, the information available on the high prevalence of HEV infections in swine (Meng et al. 1999; Pina et al. 2000) strongly suggests that HEV strains of swine origin transmitted through direct or indirect contact with infected pigs or through contaminated swine products or manure contribute to the circulation of HEV genotype 3 in the population.

The results of the present study show no detectable differences in the level of excretion of HAV in Barcelona, a place where all pre-adolescents have been vaccinated against HAV since 1999, compared to Valencia, where only risk groups are vaccinated. This observation strongly suggests that the reduction in HAV circulation does not occur in a single area of Spain, nor is this decrease related to a specific vaccination program. For HAV, 27.8% of sewage samples collected from 2000 to 2004 were positive.
This period could be considered a transition period and the results could be attributed to the ongoing improvements in sanitation. From the samples collected since 2006, HAV was detected in only one sample (3.1%) from Barcelona and in one sample (2.4%) from Valencia. Meanwhile, during this study, HAV was analysed in faecally contaminated water samples collected in north Egypt, an endemic area for this virus (Fix et al. 2006), which was present in a very high proportion of analysed samples (authors’ unpublished results). Implementation of wastewater treatments and sewage treatment plants is a determinant factor in reducing faecally transmitted diseases in a population. From 1992 to 2008, the region of Catalonia experienced a significant increase in the number of sewage treatment plants (from 91 to 343) and in the total volume of wastewater depurated (from 991,892 to 2,786,871 m³/day).

The results obtained on the prevalence of HEV showed a more stable excretion pattern in the population of NE Spain, with percentages of 28.1, 38.9 and 31.7% of positive urban sewage samples over the years studied after 2000 (Table 1). The concentration of HEV in positive urban sewage samples was lower than that of HAV, being \(10^5\) CG/ml and \(10^4\) CG/ml, respectively. The observed concentration of HAdV was much higher, which registered more than \(10^3\) CG/ml in all the samples analysed from Spain. This finding demonstrates the presence of faecal contamination in the samples studied and supports the applicability of human adenoviruses as indicators of human faecal contamination.

The studies of the amplified viral sequences showed the variability of HAV and HEV strains present in sewage. Although the sizes of the sequences were small, previous comparative studies have shown that the sequences analysed can be used for typification studies. In addition, the phylogenetic trees produced in this study show equivalent topographies to those obtained when longer sequences were analysed (Pina et al. 2001; Clemente-Casares et al. 2003; Bofill-Mas et al. 2006). All the viral sequences obtained differed from the positive controls used in the experiments, thereby demonstrating the absence of cross contamination in the study. All the HEV strains identified were classified as genotype 3, except one sample which was in genotype 1. Furthermore, the genotyping of the viruses identified in clinical serum samples revealed the same results as for HEV, that is to say, most cases showed the presence of genotype 3, while one sample belonged to genotype 1, in this case in a patient who had recently travelled to India. The variability of HEV sequences detected in urban sewage samples in the studied area shows that diverse genotype 3 strains simultaneously circulate in the population. The sporadic detection of HEV genotype 1, considered typical of endemic regions and without animal reservoirs identified to date, indicates that HEV infections are well established in the human population in Europe and that a small proportion of genotype 1 strains also circulate in industrialized countries.

The results obtained in this study strongly suggest that HEV has replaced HAV as the most frequently detected hepatitis virus potentially transmitted through local faecally contaminated water or food in eastern Spain.

In industrialized countries such as the United States, a decline in the frequency of hepatitis A and in the mortality rate was observed in the past decade (serologic results and other approach) (Bell et al. 2005; Vogt et al. 2008). The study of the viruses present in urban sewage provides a useful approach to obtaining global information on their molecular epidemiology considering also subclinical infections in the population (Pina et al. 2001).

The dramatic reduction in the presence of HAV in sewage observed in eastern Spain could be most likely related to improvements in sanitation. However, these improvements have not had an equivalent effect on the circulation of HEV genotype 3 in the area. The continued circulation of this genotype would be maintained through animal hosts as HEV infections in swine represent an external source of HEV in the human population.

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REFERENCES


Direcció general de salut pública, consellera de sanitat, Generalitat Valenciana 2008 (available at http://www.spesan.gva.es/rtn/).


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